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3 Letter to the Editor

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5 **A plea for standardized nomenclature of snake venom toxins**

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27 "...they have all one language; and this they begin to do: and now nothing will be restrained
28 from them, which they have imagined to do. Go to, let us go down, and there confound their
29 language, that they may not understand one another's speech."

30 Genesis, Chapter 11, verses 6-7

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32 In the book of Genesis a spiteful God replaces a single, unifying language with a diversity of
33 different ones, removing at a stroke mankind's ability to work together towards a common goal.
34 We believe that the Tower of Babel myth has clear relevance to the snake venom literature,
35 particularly to the nomenclature of snake venom toxins.

36 An established nomenclatural system for human genes, administered by the Human Genome
37 Organisation (HUGO) Gene Nomenclature Committee (HGNC), has existed since 1979 (Shows
38 et al., 1979) and similar systems also exist for mice (administered by the International
39 Committee on Standardized Genetic Nomenclature for Mice) and zebrafish (administered by the
40 Zebrafish Nomenclature Committee (ZNC)), as well as various other animal models. As a result,
41 orthology, paralogy and the evolutionary relationships of these genes are clear for all to see. A
42 nomenclatural committee was established by the International Society on Toxinology (IST) in
43 1992 after several exchanges in the 'Letters to the Editor' section of *Toxicon* in the early 1990's
44 (Aird, 1990; Kaiser, 1990; Kumar et al., 1990), and more recently there have been suggestions
45 for rational nomenclatural systems in spiders (King et al., 2008), scorpions (Tytgat et al., 1999),
46 sea anemones (Oliveira et al., 2012) and centipedes (Undheim et al., 2014). However, despite
47 over 20 years of asking, a nomenclatural system for reptile toxins has still not been established.
48 The lack of widely-accepted, unifying standards for the nomenclature of snake venom toxins
49 has resulted in huge diversity and disparity in the literature. In most cases the names assigned
50 provide no insight into the evolutionary origins or relationships of the toxins, complicating
51 comparative studies of venom composition, function or evolution both within and between
52 species. For example, the similar sounding *ophanin* (accession AY181984, (Yamazaki et al.,

53 2003)), *opharin* (accession AY299475, Direct submission) and *ohanin* (accession DQ103590,
54 (Pung et al., 2005)) from the king cobra (*Ophiophagus hannah*) are members of two different
55 gene families – *ophanin* and *opharin* are cysteine-rich secretory proteins (CRISPs) and *ohanin*
56 is a vespryn-like gene. Conversely, the differently sounding *pseutarin C* from *Pseudonaja textilis*
57 (accession AY260939, (Rao et al., 2004)) and *trocarin D* from *Tropidechis carinatus* (accession
58 DQ017707, (Reza et al., 2007)) are both *coagulation factor X* genes and *barietin* from *Bitis*
59 *arietans* (accession FJ554635, (Yamazaki et al., 2009)), *apiscin* from *Agkistrodon piscivorus*
60 (accession FJ554636, (Yamazaki et al., 2009)) and *cratrin* from *Crotalus atrox* (accession
61 FJ554637, (Yamazaki et al., 2009)) are all *vascular endothelial growth factor F (vegf-f)* genes.
62 The snake venom literature is replete with further examples, all of which obscure the
63 relationships and functions of the toxins themselves – who would imagine that “cobra venom
64 factor” which has been known since the late 19th century (Vogel, 1991) is part of the
65 complement system from its name alone? Indeed, we have recently shown (Hargreaves et al.,
66 2014a) that the *complement c3* gene has in fact been duplicated in cobras, giving rise to *cobra*
67 *venom factor*, which should more accurately be called *complement c3b* to reflect its true
68 evolutionary history.

69 We have previously suggested (Hargreaves et al., 2014b) that the “evolutionary characterization
70 code” proposed by the Anolis Gene Nomenclature Committee (AGNC) for lizards of that genus
71 (Kusumi et al., 2011) should be applied to snake venom toxins, and we reiterate that suggestion
72 here. Full details of this classification code can be found in the relevant reference (Kusumi et
73 al., 2011), but we provide a brief summary of some of the most relevant points below:

- 74 • Gene symbols should be written completely in lower case and in italics, for example
75 *“gene2”*.
- 76 • Punctuation (dashes, periods, slashes) should not be used unless they are part of a
77 human or mouse gene symbol.

- 78 • Whenever orthology can be assigned, this should be present in the gene name, for
79 example if the human gene symbol is “*GENE2*” then the reptile gene symbol would be
80 “*gene2*”.
- 81 • If an orthologous gene cannot be identified in any currently sequenced genome, a novel
82 name may be selected by the investigators.
- 83 • Gene symbols should not start with letters to indicate genus/species.
- 84 • Gene duplicates should be assigned the suffix “a” or “b” to indicate them as being
85 paralogs, e.g. *gene2a* and *gene2b*.

86 In addition, we would suggest that where toxins are known only from peptide or protein
87 sequences, without accompanying characterized gene sequences, they should be named on
88 the basis of similarity to existing toxins using the suffix “-like” or be acknowledged as
89 uncharacterized toxins. We should not shy away from acknowledging uncertainty, nor from
90 presenting challenges to future researchers. That being said, it seems likely that the availability
91 of genomic and transcriptomic data from an ever-growing range of species (see next section)
92 will go at least some way to facilitating the identification of toxins in proteomic studies.

93 With the recent publication of the whole genome sequences of two species of snake (the
94 Burmese python *Python molurus bivittatus* and the king cobra, *Ophiophagus hannah* (Castoe et
95 al., 2013; Vonk et al., 2013)), ongoing projects for several more (Castoe et al., 2011) and
96 increasing amounts of transcriptomic data becoming available for a wide variety of species, it is
97 now more important than ever that these data are made as easily accessible, understandable
98 and useable as possible. Whilst we appreciate the historical nature of the names of many snake
99 venom toxins and do not argue that their sometimes rich history should be neglected, we must
100 also consider the needs of the present and the future. The field of snake venom research should
101 not distance itself from the rest of biology via the continued use of non-standardized
102 nomenclature. If we want to facilitate collaboration with those from other research fields it is

103 important that we all speak the same language – only then can we work together to better
104 understand the origins and evolution of snake venom; its composition and function and its
105 possible utility in the development of novel therapeutics.

106 We note that *Toxicon*, unlike many other journals, does not currently suggest or enforce a
107 standard nomenclature of genes reported in its papers. We therefore respectfully suggest that
108 the Editors of this journal are well-placed to lead the way in the acquisition and development of
109 a standardized nomenclatural system for snake venom toxins (and, indeed, for the toxins of
110 other venomous animals). After all, what is the point of being an Editor if you can't play God
111 once in a while?

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118 References

119 Card, D.C., Schield, D.R., Reyes-Velasco, J., Adams, R.H., Mackessy, S.P., Castoe, T.A. The
120 genome of the Prairie Rattlesnake (*Crotalus viridis viridis*). Conference abstract from Biology of
121 the pitvipers symposium 2, 4th-7th June 2014. Tulsa, OK, USA.

122 Aird, S.D., 1990. Call for an IST nomenclatural committee. *Toxicon* 28, 136-137.

123 Castoe, T.A., Bronikowski, A.M., Brodie, E.D.,3rd, Edwards, S.V., Pfrender, M.E., Shapiro,
124 M.D., Pollock, D.D., Warren, W.C., 2011. A proposal to sequence the genome of a garter snake
125 (*Thamnophis sirtalis*). *Stand. Genomic Sci.* 4, 257-270.

126 Castoe, T.A., de Koning, A.P., Hall, K.T., Card, D.C., Schield, D.R., Fujita, M.K., Ruggiero, R.P.,
127 Degner, J.F., Daza, J.M., Gu, W., Reyes-Velasco, J., Shaney, K.J., Castoe, J.M., Fox, S.E.,
128 Poole, A.W., Polanco, D., Dobry, J., Vandewege, M.W., Li, Q., Schott, R.K., Kapusta, A., Minx,
129 P., Feschotte, C., Uetz, P., Ray, D.A., Hoffmann, F.G., Bogden, R., Smith, E.N., Chang, B.S.,
130 Vonk, F.J., Casewell, N.R., Henkel, C.V., Richardson, M.K., Mackessy, S.P., Bronikowski, A.M.,
131 Yandell, M., Warren, W.C., Secor, S.M., Pollock, D.D., 2013. The Burmese python genome
132 reveals the molecular basis for extreme adaptation in snakes. *Proc. Natl. Acad. Sci. U. S. A.*
133 110, 20645-20650.

134 Hargreaves, A.D., Swain, M.T., Hegarty, M.J., Logan, D.W. and Mulley, J.F., 2014a Restriction
135 and recruitment – gene duplication and the origin and evolution of snake venom toxins. *Genome*
136 *Biol. Evol.* 6, 2088-2095.

137 Hargreaves, A.D., Swain, M.T., Logan, D.W., Mulley, J.F., 2014b. Testing the Toxicofera:
138 comparative reptile transcriptomics casts doubt on the single, early evolution of the reptile
139 venom system. bioRxiv doi: <http://dx.doi.org/10.1101/006031>

140 Kaiser, I.I., 1990. Toxin nomenclature. *Toxicon* 28, 137-138.

141 King, G.F., Gentz, M.C., Escoubas, P., Nicholson, G.M., 2008. A rational nomenclature for
142 naming peptide toxins from spiders and other venomous animals. *Toxicon* 52, 264-276.

143 Kumar, T., Radha, R., Rambhav, S., 1990. What's in a name? *Toxicon* 28, 135-136.

144 Kusumi, K., Kulathinal, R.J., Abzhanov, A., Boissinot, S., Crawford, N.G., Faircloth, B.C., Glenn,
145 T.C., Janes, D.E., Losos, J.B., Menke, D.B., Poe, S., Sanger, T.J., Schneider, C.J., Stapley, J.,
146 Wade, J., Wilson-Rawls, J., 2011. Developing a community-based genetic nomenclature for
147 anole lizards. *BMC Genomics* 12, 554-2164-12-554.

148 Oliveira, J.S., Fuentes-Silva, D., King, G.F., 2012. Development of a rational nomenclature for
149 naming peptide and protein toxins from sea anemones. *Toxicon* 60, 539-550.

150 Pung, Y.F., Wong, P.T., Kumar, P.P., Hodgson, W.C., Kini, R.M., 2005. Ohanin, a novel protein
151 from king cobra venom, induces hypolocomotion and hyperalgesia in mice. *J. Biol. Chem.* 280,
152 13137-13147.

153 Rao, V.S., Swarup, S., Kini, R.M., 2004. The catalytic subunit of pseutarin C, a group C
154 prothrombin activator from the venom of *Pseudonaja textilis*, is structurally similar to mammalian
155 blood coagulation factor Xa. *Thromb. Haemost.* 92, 509-521.

156 Reza, M., Swarup, S., Kini, R., 2007. Structure of two genes encoding parallel prothrombin
157 activators in *Tropidechis carinatus* snake: gene duplication and recruitment of factor X gene to
158 the venom gland. *J. Thromb. Haemost.* 5, 117-126.

159 Shows, T., Alper, C., Bootsma, D., Dorf, M., Douglas, T., Huisman, T., Kit, S., Klinger, H.,
160 Kozak, C., Lalley, P., 1979. International system for human gene nomenclature (1979) ISGN
161 (1979). *Cytogenet. Genome Res.* 25, 96-116.

162 Tytgat, J., Chandy, K.G., Garcia, M.L., Gutman, G.A., Martin-Eauclaire, M., van der Walt, Jurg
163 J, Possani, L.D., 1999. A unified nomenclature for short-chain peptides isolated from scorpion
164 venoms: α -KTx molecular subfamilies. *Trends Pharmacol. Sci.* 20, 444-447.

165 Undheim, E.A.B, Jones, A., Clauser, K.R., Holland, J.W., Pineda, S.S., King, G.F., Fry, B.G.
166 2014. Clawing through Evolution: Toxin Diversification and Convergence in the Ancient Lineage
167 Chilopoda (Centipedes). *Mol Biol Evol* 31, 2124-2148.

168 Vonk, F.J., Casewell, N.R., Henkel, C.V., Heimberg, A.M., Jansen, H.J., McCleary, R.J.,
169 Kerckamp, H.M., Vos, R.A., Guerreiro, I., Calvete, J.J., Wuster, W., Woods, A.E., Logan, J.M.,

170 Harrison, R.A., Castoe, T.A., de Koning, A.P., Pollock, D.D., Yandell, M., Calderon, D., Renjifo,
171 C., Currier, R.B., Salgado, D., Pla, D., Sanz, L., Hyder, A.S., Ribeiro, J.M., Arntzen, J.W., van
172 den Thillart, G.E., Boetzer, M., Pirovano, W., Dirks, R.P., Spaink, H.P., Duboule, D., McGlinn,
173 E., Kini, R.M., Richardson, M.K., 2013. The king cobra genome reveals dynamic gene evolution
174 and adaptation in the snake venom system. *Proc. Natl. Acad. Sci. U. S. A.* 110, 20651-20656.

175 Yamazaki, Y., Hyodo, F., Morita, T., 2003. Wide distribution of cysteine-rich secretory proteins
176 in snake venoms: isolation and cloning of novel snake venom cysteine-rich secretory proteins.
177 *Arch. Biochem. Biophys.* 412, 133-141.

178 Yamazaki, Y., Matsunaga, Y., Tokunaga, Y., Obayashi, S., Saito, M., Morita, T., 2009. Snake
179 venom Vascular Endothelial Growth Factors (VEGF-Fs) exclusively vary their structures and
180 functions among species. *J. Biol. Chem.* 284, 9885-9891.